# **WENDER GUIDE CORP. G1802**

# **PEI 40K Transfection Reagent**

Product #: G1802

Quantity: 5 x 1 mL

### **Product Description**

- Plasmid DNA transfection suitable for cells, with low cytotoxicity, high transfection efficiency, and low cost.
- This transfection reagent contains polyethyleneimine (PEI) covalently bound to human transferrin, providing a highly specific DNA transformation method known as transferrinfection.
- Transferrinfection is a high-efficiency nucleic acid delivery system based on transferrin receptor-mediated endocytosis. In the PEI-transferrinfection system, the gene transfer efficiency of PEI/DNA complexes are combined with the specific mechanism of receptormediated endocytosis via the transferrin receptor.
  - PEI mediates efficient gene transfer into a variety of cells.
  - $\circ$  30 1,000-fold enhanced transfection efficiency depending on the cell line.
  - o Gentle DNA transfection method and employs physiological uptake mechanisms.
  - Transfection efficacy depends on the cell type, the level of surface transferrin receptor expression.
    - Very high transfection rates have been shown for the tested tumor cell lines B16F10 melanoma, Neuro 2A neuroblastoma, and a variety of primary human melanoma cell lines.
    - In other established cell lines, such as HeLa, CHO, Jurkat, K562, HepG2, and COS, the PEI-Transferrinfection works with high efficiency, excellent reproducibility and with the advantage of being an extremely gentle procedure.
- The concentration of PEI 40K, the main component of this product, is 1 mg/ml.

### Storage

- Store at -20<sup>o</sup>C for long-term storage (avoid repeated freeze-thaw cycles).
- $\circ$  Can be stored at 4<sup>o</sup>C for up to 3 months.



## Protocol

- 1. Preparation of adherent cells (protocol provided for one-well of 6-well plate; refer to table below for other vessel types):
  - A. The day before transfection, plate cells at an appropriate density so 70 85% confluent on the day of transfection.
  - B. Before transfection, remove media and wash cells with PBS before replacing with 1.8 mL of Opti-MEM (can also use basal medium without serum or low serum (below 5%)).
  - C. Placed the cells back in the incubator and proceed with preparation of transfection complexes (Step 3).
- 2. Preparation of suspension cells:
  - A. On the day of transfection, centrifuge an appropriate number of cells to remove the medium and resuspended with Opti-MEM (can also use basal medium without serum or low serum (below 5%)).
  - B. Placed the cells back in the incubator and proceed with preparation of transfection complexes (Step 3).
- 3. Transfection complexes were prepared:
  - A. Preparation of **Solution A**: 100  $\mu$ L of Opti-Mem (or serum-free base medium) and 2  $\mu$ g of plasmid DNA. Mix well while injecting air into solution using pipette.
  - B. Preparation of **Solution B**: 100  $\mu$ L of Opti-Mem (or serum-free medium) and 6-8  $\mu$ L of PEI 40 K Transfection reagent. Mix well while injecting air into solution using pipette.
- 4. Slowly add **Solution B** to **Solution A.** Mix well while injecting air into solution using pipette.
- 5. Incubate mixed solution at room temperature for 15 min to form DNA-PEI transfection complex.



- 6. Cell transfection: add the transfection complex to the cells dropwise while distributing evenly. Swivel the cell culture flask to ensure even distribution of the cell transfection reagent.
- 7. Place the cells in the incubator.
- 8. After 4 6 h, replace with complete media.

| Vessel Type        | Growth<br>area<br>(cm²) | Number of inoculated cells | DNA content<br>(μg) | ΡΕΙ<br>(μL) | Transfection<br>complex<br>volume<br>(μL) | Total volume<br>(mL) |
|--------------------|-------------------------|----------------------------|---------------------|-------------|---|----------------------|
| 96-well plates     | 0.3                     | (2-4)×10 <sup>4</sup>      | 0.1                 | 0.3-0.4     | 10  | 0.1                  |
| 24-well plates     | 1.9                     | (1.2-2.4)×10 <sup>5</sup>  | 0.5-1               | 1.5-4       | 50  | 0.5                  |
| 12-well plates     | 3.8                     | (2.4-4.8)×10 <sup>5</sup>  | 1-2                 | 3-8         | 100                                       | 1                    |
| 6-well plates      | 9.5                     | (6-10)×10 <sup>5</sup>     | 2-4                 | 6-16        | 200                                       | 2                    |
| 10 cm culture dish | 55                      | (4-6)×10 <sup>6</sup>      | 12-24               | 36-96       | 1000                                      | 12                   |
| T75 culture flask  | 75                      | (6-10)×10 <sup>6</sup>     | 18-36               | 54-144      | 1000                                      | 15                   |

Note: amount of DNA, PEI, and cell numbers need to be optimized for best results.

NOTES:

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